Effect of PON1 on dichlorvos toxicokinetics

Na-Na Wang, Li Yuan, Heng Dai, Zhen-Kun Han, Min Zhao

ABSTRACT

Objectives To provide toxicokinetic and clinical evidence of the hydrolytic effect of paraoxonase-1 (PON1) on acute organophosphate poisoning in rats.

Methods 40 male Wistar rats were randomised into four equal groups. Dichlorvos administration group (A group) underwent dichlorvos injection (dissolved in corn oil) using intraperitoneal (ip) dose of 10 mg/kg. PON1 pretreatment group (B group) was injected with PON1 in the tail vein (intravenous), dose 9600 U/kg, 30 min prior to dichlorvos administration. In the treatment group (C group), atropine 0.05 mg/kg and pyraloxime chloride (PAM-CI) 120 mg/kg were injected intravenously within 2 min after dichlorvos administration. Finally, in the co-treatment group (D group), PON1 was injected intravenously with a dose of 9000 U/kg 30 min prior to dichlorvos administration; atropine 0.05 mg/kg and PAM-CI 120 mg/kg were injected intravenously within 2 min after dichlorvos administration. Blood was collected after administration. Plasma dichlorvos concentration was detected by liquid chromatography-mass spectra (LC-MS) method and clinical signs were observed. Toxicokinetic parameters were calculated in a statistical moment model.

Results AUC (0→∞) in group B was statistically different from that in groups A and C (p<0.05), while it was not different from group D (p>0.05); there was no statistical difference between group A and group C (p>0.05). The statistical results of Cmax were the same as those of AUC (0→∞). There were no differences of MRT between four groups (p>0.05). Clinical signs can be improved by PON1 and atropine + PAM-CI, and co-treatment can relieve signs more effectively.

Conclusion PON1 can decrease the amount of dichlorvos that entered the blood, lowered the peak concentration and relieved clinical signs.

Serum paraoxonase-1 (PON1) is an A-esterase that is associated with high-density lipoprotein (HDLs). The products of PON1 gene exist widely in mammals, with the highest activity in liver and blood. It is involved in the detoxification of organophosphate insecticides, such as chlorpyrifos oxon, diazoxon, paraoxon and dichlorvos. As a result, it is considered to have great significance in the detoxification of organophosphate compounds. Therefore, PON1 may prevent tissue damage from organophosphate toxicity, especially in the central nervous system. There have been studies designed to evaluate the protective effect of PON1 to organophosphates in vitro or by measuring inhibition of acetyl cholinesterase in different tissues, but studies on organophosphate concentration changes in vivo are rare. In the present study, purified rabbit serum PON1 was administered to rats intravenously and concentration of dichlorvos in blood was detected at different time points to analyse the effect of PON1 on toxicokinetics of dichlorvos.

MATERIALS AND METHODS

Material

Dichlorvos (>99.5% pure) was obtained from Tianjin Agriculture Co., Ltd; acetonitrile, indomethacin and methanoic acid (chromatographic pure) were homemade reagents. Clean grade male Wistar rats, weighing 250–500 g, were obtained from Beijing Vital River Experimental Animal Co., Ltd. The animals were fed ad libitum in the experiment.

AKTA purifier automatic chromatography was manufactured by GE Company (Bridgeport, Connecticut, USA). TSQ Quantum Ultra, produced by Thermo Finnigan (San Jose, California, USA). Eppendorf 5417-R refrigerated centrifuge was manufactured by Eppendorf Company (Hamburg, Germany).

Dose schedule and sample collection

40 male Wistar rats were randomised into four equal groups. The dichlorvos administration group (A group) underwent dichlorvos injection (dissolved in corn oil) using intraperitoneal (ip) dose of 10 mg/kg. The PON1 pretreatment group (B group) was injected with PON1 in the tail vein (intravenous), dose 9600 U/kg, 30 min prior to dichlorvos administration (10 mg/kg ip). In the treatment group (C group), atropine 0.05 mg/kg and pyraloxime chloride (PAM-CI) 120 mg/kg were injected intravenously within 2 min after dichlorvos administration (10 mg/kg ip). In the co-treatment group (D group), PON1 was injected intravenously in a dose of 9000 U/kg 30 min prior to dichlorvos administration (10 mg/kg ip); atropine 0.05 mg/kg and PAM-CI 120 mg/kg were injected intravenously within 2 min after dichlorvos administration.

Blood of 0.2 ml was collected at 3 min, 5 min, 10 min, 20 min, 30 min, 1 h, 2 h, 4 h and 6 h after administration from the eye veins. Blood was centrifuged at 5000 r for 5 min and the supernatant was collected and stored in –80°C for dichlorvos detection.

Clinical signs after dichlorvos poisoning were observed. According to Gaidukov,6 all clinical signs noted following dichlorvos intoxication were categorised to mild, moderate or severe reactions. Mild reactions were characterised by straub tail and/or ataxia and/or diarrhoea. Moderate reactions consisted of, in addition, decreased motor activity and/or dyspnoea, while animals with severe reactions exhibited ventral position and/or tremors as well. The overall reactions observed following dichlorvos intoxication were scored using semi-quantitative grading of five grades (0–4), taking into
consideration the severity of the reactions (0 = no reactions, 1 = mild reactions, 2 = moderate reactions, 3 = severe reactions, 4 = mortality). Because there were no deaths in the experiment, no rats scored 4.

**Purification of rabbit serum PON1**

Paraoxonase was purified from rabbit serum as previously described in detail. The collected protein was subjected to centrifugal ultrafiltration on centrulps YM-10 (Millipore, Billerica, Massachusetts, USA) so as to concentrate to >5000 U/ml. The enzyme was dissolved in buffer solution (156 mM NaCl, 0.1 mM CaCl2) for injection before usage.

**LC-MS method**

**Sample preparation**

Plasma aliquots of 50 μl was added to 2.5 μl of the internal standard indomethacin solutions that was prepared in methanol of 50 ng/ml. Then, acetonitrile (100 μl) was added to the mixture. The resulting mixture was vortex-mixed for 1 min and then centrifuged at 12000 r for 5 min. The supernatant was filtered through a 0.45-μm Millex®-LH filter, and 20 μl of the filtrate was injected into the LC-APCI-MS.

**LC-MS condition**

The HPLC analysis was performed on a Diamonsil C18 column (150×4.6 mm inner diameter, 5 μm) equipped with an phenomenex C18 guard column (4×3.0 mm inner diameter) at room temperature. Fluid phase was used with solvent consisting of acetonitrile–water–methanoic acid (90:10:0.2, v/v/v) at a flow rate of 0.45 ml/min. The solutions were filtered through a 0.45 μm Millex®-LH filter before use.

A Thermo Finnigan TSQ Quantum Ultra tandem mass spectrometer equipped with an APCI interface operated in the positive ion mode. The capillary voltage was set to 4.2 kV. The capillary temperature was 270°C, and the desolvation temperature was 420°C. Nitrogen was used as desolvation gas (pressure 30 Arb) and assist gas (pressure 5 Arb), and argon was used as collision gas at a pressure of 1.2 mTorr. Detection was performed in selected reaction monitoring (SRM) mode. The collision-induced dissociation voltages were 23 eV (dichlorvos) and 44 eV (indomethacin), respectively. Ionic reaction for quantitative analysis was m/z 221 → m/z 109 (dichlorvos) and m/z 358 → m/z 111 (indomethacin), respectively. Scan time was 0.3 s.

Following analysis by an LC-APCI-MS system, the retention time for dichlorvos was 4.07 s, and that for indomethacin was 4.92 s (figure 1).

**Linearity**

Added dichlorvos standards to blank plasma detected the ratio of peak area of 6 μg/ml, 5 μg/ml, 2 μg/ml, 1 μg/ml, 0.5 μg/ml and 0.2 μg/ml of dichlorvos to internal standard peak area.

With defined dichlorvos standards peak area as X-axis, ratio of dichlorvos standards peak area to internal standard peak area as Y-axis, we got the regression line of dichlorvos: \( Y = -0.0404282 + 0.16048X \), \( R^2 = 0.9945 \).

The linearity was 0.2–6 μg/ml.

**Statistics**

A statistical moment model was used to calculate toxicokinetics parameters for dichlorvos by spss7 software. Statistical analyses were conducted using SPSS16.0 software.

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**RESULTS**

**Toxicokinetics of dichlorvos**

**Dichlorvos concentration**

The concentrations of dichlorvos in B group were statistically different from A group (p<0.05), while atropine + PAM-CI did not alter dichlorvos concentration statistically (p>0.05) (table 1).

**Dichlorvos toxicokinetics parameters**

The toxicokinetics parameter differences between four groups were performed by ANOVA. There were no statistical differences between AUC (0→∞) of groups A and C, but the effect of PON1 was obvious in groups B and D when compared with group A. There were no statistical differences between MRT (0→∞) of the four groups. Cmax was obtained by observing the peak concentrations in table 1. The statistical result coincided with that of AUC (0→∞) (table 2).

**Clinical signs**

The clinical signs were alleviated by PON1, atropine + PAM-CI and co-treatment, and the effect of PON1 was not different with atropine + PAM-CI statistically. However, co-treatment achieved the best effect (table 3).

**DISCUSSION**

The 1/2 LD50 (LD50, 18–20 mg/kg by ip) of dichlorvos was selected so that the most severe signs of poisoning would be elicited but no death arose through preliminary experiment. The

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**Table 1**

<table>
<thead>
<tr>
<th>Time (h)</th>
<th>A group (μg/ml)</th>
<th>B group (μg/ml)</th>
<th>C group (μg/ml)</th>
<th>D group (μg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.05</td>
<td>1.84±0.85</td>
<td>0.76±0.17</td>
<td>1.83±0.40</td>
<td>0.76±0.07</td>
</tr>
<tr>
<td>0.08</td>
<td>3.31±0.31</td>
<td>1.22±0.13</td>
<td>3.13±0.27</td>
<td>1.33±0.21</td>
</tr>
<tr>
<td>0.17</td>
<td>5.36±0.63</td>
<td>2.91±0.18</td>
<td>5.28±0.55</td>
<td>2.90±0.32</td>
</tr>
<tr>
<td>0.33</td>
<td>2.85±0.30</td>
<td>1.57±0.26</td>
<td>2.74±0.42</td>
<td>1.54±0.05</td>
</tr>
<tr>
<td>0.5</td>
<td>1.93±0.26</td>
<td>0.99±0.18</td>
<td>1.87±0.34</td>
<td>0.95±0.17</td>
</tr>
<tr>
<td>1</td>
<td>1.35±0.22</td>
<td>0.64±0.08</td>
<td>1.25±0.10</td>
<td>0.64±0.10</td>
</tr>
<tr>
<td>2</td>
<td>1.09±0.21</td>
<td>0.35±0.07</td>
<td>1.05±0.16</td>
<td>0.35±0.01</td>
</tr>
<tr>
<td>4</td>
<td>0.66±0.16</td>
<td>0.23±0.01</td>
<td>0.65±0.09</td>
<td>0.23±0.03</td>
</tr>
<tr>
<td>6</td>
<td>0.37±0.06</td>
<td>0.22±0.01</td>
<td>0.35±0.02</td>
<td>0.21±0.01</td>
</tr>
</tbody>
</table>

All the data were expressed as mean±SD. *p<0.01, concentration data were compared with A group by t test. There were statistical differences between group B and group A and between group C and group A.
Table 2  Toxicokinetics parameters of dichlorvos

<table>
<thead>
<tr>
<th>Parameter</th>
<th>A group</th>
<th>B group</th>
<th>C group</th>
<th>D group</th>
</tr>
</thead>
<tbody>
<tr>
<td>AUC(0 → ∞) (g/ml*h)</td>
<td>11.24±1.63</td>
<td>4.25±0.41</td>
<td>10.71±0.97</td>
<td>4.67±0.99</td>
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<tr>
<td>MRT (h)</td>
<td>7.93±1.40</td>
<td>8.50±2.05</td>
<td>7.65±0.87</td>
<td>13.89±5.80</td>
</tr>
<tr>
<td>Cmax (μg/ml)</td>
<td>5.36±0.63</td>
<td>2.91±0.18</td>
<td>5.28±0.55</td>
<td>2.90±0.32</td>
</tr>
</tbody>
</table>

All the data were expressed as mean± SD.

Table 3  Clinical signs scores of four groups

<table>
<thead>
<tr>
<th>Group</th>
<th>Score</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0</td>
</tr>
<tr>
<td>A group</td>
<td>0</td>
</tr>
<tr>
<td>B group</td>
<td>0</td>
</tr>
<tr>
<td>C group</td>
<td>0</td>
</tr>
<tr>
<td>D group</td>
<td>2</td>
</tr>
</tbody>
</table>

Ridit analysis was used to analyse the differences between the data of clinical signs scores of four groups,’ where p<0.05 was regarded as significant.

REFERENCES
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