Effect of PON1 on dichlorvos toxicokinetics

Na-Na Wang, Li Yuan, Heng Dai, Zhen-Kun Han, Min Zhao

ABSTRACT

Objectives To provide toxicokinetic and clinical evidence of the hydrolytic effect of paraoxonase-1 (PON1) on acute organophosphate poisoning in rats.

Methods 40 male Wistar rats were randomised into four equal groups. Dichlorvos administration group (A group) underwent dichlorvos injection (dissolved in corn oil) using intraperitoneal (ip) dose of 10 mg/kg. PON1 pretreatment group (B group) was injected with PON1 in the tail vein (intravenous), dose 9600 U/kg, 30 min prior to dichlorvos administration. In the treatment group (C group), atropine 0.05 mg/kg and pyr nonsense chloride (PAM-CI) 120 mg/kg were injected intravenously within 2 min after dichlorvos administration. Finally, in the co-treatment group (D group), PON1 was injected intravenously with a dose of 9000 U/kg 30 min prior to dichlorvos administration; atropine 0.05 mg/kg and PAM-CI 120 mg/kg were injected intravenously within 2 min after dichlorvos administration. Blood was collected after administration. Plasma dichlorvos concentration was detected by liquid chromatography-mass spectra (LC-MS) method and clinical signs were observed. Toxicokinetic parameters were calculated in a statistical moment model.

Results AUC (0→∞) in group B was statistically different from that in groups A and C (p<0.05), while it was not different from group D (p>0.05); there was no statistical difference between group A and group C (p>0.05). The statistical results of Cmax were the same as those of AUC (0→∞). There were no differences of MRT between four groups (p>0.05). Clinical signs can be improved by PON1 and atropine + PAM-CI, and co-treatment can relieve signs more effectively.

Conclusion PON1 can decrease the amount of dichlorvos that entered the blood, lowered the peak concentration and relieved clinical signs.

Serum paraoxonase-1 (PON1) is an A-esterase that is associated with high-density lipoprotein (HDLs). The products of PON1 gene exist widely in mammals, with the highest activity in liver and blood. It is involved in the detoxification of organo- phosphor pesticides, such as chlorpyrifos oxon, diazoxon, paraoxon and dichlorvos.1, 2 As a result, it is considered to have great significance in the detoxification of organophosphate compounds. Therefore, PON1 may prevent tissue damage from organophosphate toxicity, especially in the central nervous system. There have been studies designed to evaluate the protective effect of PON1 to organophosphates in vitro or by measuring inhibition of acetyl cholinesterase in different tissues, but studies on organo- phosphor concentration changes in vivo are rare.3-5 In the present study, purified rabbit serum PON1 was administered to rats intravenously and concentration of dichlorvos in blood was detected at different time points to analyse the effect of PON1 on toxicokinetics of dichlorvos.

MATERIALS AND METHODS

Material

Dichlorvos (>99.5% pure) was obtained from Tianjin Agriculture Co., Ltd; acetonitrile, indomethacin and methanoic acid (chromatographic pure) were homemade reagents.

Clean grade male Wistar rats, weighing 250–500 g, were obtained from Beijing Vital River Experimental Animal Co., Ltd. The animals were fed ad libitum in the experiment.

ÅKTA purifier automatic chromatography was manufactured by GE Company (Bridgeport, Connecticut, USA). TSQ Quantum Ultra, produced by Thermo Finnigan (San Jose, California, USA). Eppendorf 5417-R refrigerated centrifuge was manufactured by Eppendorf Company (Hamburg, Germany).

Dose schedule and sample collection

40 male Wistar rats were randomised into four equal groups. The dichlorvos administration group (A group) underwent dichlorvos injection (dissolved in corn oil) using intraperitoneal (ip) dose of 10 mg/kg. The PON1 pretreatment group (B group) was injected with PON1 in the tail vein (intravenous), dose 9600 U/kg, 30 min prior to dichlorvos administration (10 mg/kg ip). In the treatment group (C group), atropine 0.05 mg/kg and pyr nonsense chloride (PAM-CI) 120 mg/kg were injected intravenously within 2 min after dichlorvos administration. Finally, in the co-treatment group (D group), PON1 was injected intravenously in a dose of 9000 U/kg 30 min prior to dichlorvos administration (10 mg/kg ip); atropine 0.05 mg/kg and PAM-CI 120 mg/kg were injected intravenously within 2 min after dichlorvos administration. Blood of 0.2 ml was collected at 3 min, 5 min, 10 min, 20 min, 30 min, 1 h, 2 h, 4 h and 6 h after administration from the eye veins. Blood was centrifuged at 5000 r for 5 min and the supernatant was collected and stored in −80°C for dichlorvos detection.

Clinical signs after dichlorvos poisoning were observed. According to Gaidukov,6 all clinical signs noted following dichlorvos intoxication were categorised to mild, moderate or severe reactions. Mild reactions were characterised by straub tail and/or ataxia and/or diarrhoea. Moderate reactions consisted of, in addition, decreased motor activity and/or dyspnoea, while animals with severe reactions exhibited ventral position and/or tremors as well. The overall reactions observed following dichlorvos intoxication were scored using semi-quantitative grading of five grades (0–4), taking into

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consideration the severity of the reactions (0: no reactions, 
1: mild reactions, 2: moderate reactions, 3: severe reactions, 
4: mortality). Because there were no deaths in the experiment, 
no rats scored 4.

**Purification of rabbit serum PON1**

Paraoxonase was purified from rabbit serum as previously 
described in detail. The collected protein was subjected to 
centrifugal ultrafiltration on centrulps YM-10 (Millipore, Bill- 
erica, Massachusetts, USA) so as to concentrate to >5000 U/ml. 
The enzyme was dissolved in buffer solution (156 mM NaCl, 
0.1 mM CaCl2) for injection before usage.

**LC-MS method**

**Sample preparation**

Plasma aliquot of 50 μl was added to 2.5 μl of the internal 
standard indomethacin solutions that was prepared in methanol 
of 50 ng/ml. Then, acetonitrile (100 μl) was added to the 
mixture. The resulting mixture was vortex-mixed for 1 min and 
then centrifuged at 12000 r for 5 min. The supernatant 
was filtered through a 0.45-μm Millex®-LH filter, and 20 μl of 
the filtrate was injected into the LC-APCI-MS.

**LC-MS condition**

The HPLC analysis was performed on a Diamonsil C18 column 
(150×4.6 mm inside diameter, 5 μm.) equipped with a 
phenomenex C18 guard column (4×3.0 mm inside diameter) at 
room temperature. Fluid phase was used with solvent consisting 
of acetonitrile—water—methalcoic acid (90:10:0.2, v/v/v) at 
a flow rate of 0.45 ml/min. The solutions were filtered through 
a 0.45 μm Millex®-LH filter before use.

A Thermo Finnigan TSQ Quantum Ultra tandem mass 
spectrometer equipped with an APCI interface operated in the 
positive ion mode. The capillary temperature was 270°C, and the desolvation 
temperature was 420°C. Nitrogen was used as desolavation gas (pressure 
30 Arb) and assist gas (pressure 5 Arb), and argon was used as 
collision gas at a pressure of 1.2 mTorr. Detection was 
performed in selected reaction monitoring (SRM) mode. The 
collision-induced dissociation voltages were 23 eV (dichlorvos) 
and 44 eV (indomethacin), respectively. Ionic reaction for 
quantitative analysis was m/z 221 → m/z 109 (dichlorvos) and 
m/z 358 → m/z 111 (indomethacin), respectively. Scan time 
was 0.3 s.8

Following analysis by an LC-APCI-MS system, the retention 
time for dichlorvos was 4.07 s, and that for indomethacin was 
4.92 s (figure 1).

**Linearity**

Added dichlorvos standards to blank plasma detected the ratio of 
peak area of 6 μg/ml, 5 μg/ml, 2 μg/ml, 1 μg/ml, 0.5 μg/ml and 
0.2 μg/ml of dichlorvos to internal standard peak area. 
With defined dichlorvos standards peak area as X-axis, ratio of 
dichlorvos standards peak area to internal standard peak area as 
Y-axis, we got the regression line of dichlorvos: \( Y = -0.0404282 +0.16048X \), \( R^2 = 0.9945 \). 
The linearity was 0.2–6 μg/ml.

**Statistics**

A statistical moment model was used to calculate toxicokinetics 
parameters for dichlorvos by spgg7 software. Statistical analyses 
were conducted using SPSS16.0 software.

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**RESULTS**

**Toxicokinetics of dichlorvos**

**Dichlorvos concentration**

The concentrations of dichlorvos in B group were statistically 
different from A group (p<0.05), while atropine + PAM-CI did not 
alter dichlorvos concentration statistically (p>0.05) (table 1).

**Dichlorvos toxicokinetics parameters**

The toxicokinetics parameter differences between four groups 
were performed by ANOVA. There were no statistical differences 
between AUC (0→∞) of groups A and C, but the effect of 
PON1 was obvious in groups B and D when compared with 
group A. There were no statistical differences between MRT 
(0→∞) of the four groups. Cmax was obtained by observing the 
peak concentrations in table 1. The statistical result coincided 
with that of AUC (0→∞) (table 2).

**Clinical signs**

The clinical signs were alleviated by PON1, atropine+ PAM-CI 
and co-treatment, and the effect of PON1 was not different 
with atropine+ PAM-CI statistically. However, co-treatment 
achieved the best effect (table 3).

**DISCUSSION**

The 1/2 LD₅₀ (LD₅₀, 18–20 mg/kg by ip) of dichlorvos was 
selected so that the most severe signs of poisoning would be 
elicted but no death arose through preliminary experiment. The
dose is different from the larger dose exposures that occur in clinical situations and experimentally in vitro; however, it allow us to evaluate toxicokinetics of dichlorvos.

The results of the study suggested that PON1 can effectively decrease blood dichlorvos concentration, reduce the peak concentration of dichlorvos and lessen the amount that enters the blood. It is possible that a high degree of protection was provided by PON1 pretreatment in animals challenged with organophosphates by measuring cholinesterase activities in different tissues.10–12 Our results provide further direct evidence by showing changes in concentration with PON1 pre-treatment. We also compared the hydrolytic effect of PON1 with atropine +PAM-CI; there- fore, it is postulated that co-treatment may be feasible in the clinical treatment of human organophosphate-related toxicity.

We also investigated the clinical signs of the four groups. Decreased blood dichlorvos concentration in the PON1 pretreatment group resulted in better clinical outcome when compared with the dichlorvos administration group. At this dose, the PON1 effect is comparable to that of atropine + PAM-CI, which is the most common antitodal treatment used clinically. In addition, another novel finding in our study was that the clinical signs were relieved the most by co-treatment. This may have important implications on the future treatment of organophosphates poisoning.

Interest in PON1 arises from the hypothesis that individuals with low serum activity of this enzyme would be expected to have a diminished ability to metabolise organophosphates. This hypothesis implies that serum PON1 has a pivotal role in the detoxification of the organophosphate. After the hypothesis was raised, some studies have been designed to evaluate the protective effect of PON1 by pretreating animals with PON1 before organophosphates administration. Such studies are needed before any definite inference can be drawn on the role that serum paraoxonase levels have in the protective effect on insecticides. However, in the treatment of human organophos-

<table>
<thead>
<tr>
<th>Parameter</th>
<th>A group</th>
<th>B group</th>
<th>C group</th>
<th>D group</th>
</tr>
</thead>
<tbody>
<tr>
<td>AUC(0 -&gt; infinity) (μg·mL⁻¹·h⁻¹)</td>
<td>11.24 ± 1.63</td>
<td>4.25 ± 0.41</td>
<td>10.71 ± 0.97</td>
<td>4.67 ± 0.99</td>
</tr>
<tr>
<td>MRT (h)</td>
<td>7.93 ± 1.40</td>
<td>8.50 ± 2.05</td>
<td>7.65 ± 0.87</td>
<td>13.89 ± 5.80</td>
</tr>
<tr>
<td>Cmax (μg/mL)</td>
<td>5.36 ± 0.63</td>
<td>2.91 ± 0.18</td>
<td>5.28 ± 0.55</td>
<td>2.90 ± 0.32</td>
</tr>
</tbody>
</table>

All the data were expressed as mean ± SD.

phate-related toxicity, antidotes are administered after exposure, so further investigations of PON1 should be conducted with PON1 administration after organophosphate exposure on the theoretical basis of protective function.

In conclusion, PON1 can hydrolyse organophosphates in vivo and improve clinical signs of organophosphate toxicity. The use of PON1 in organophosphates poisoning requires further investigations.

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