venous thromboembolic disease has an estimated annual incidence in developed countries of 1 in 1000 people.\textsuperscript{1} The clinical diagnosis of deep vein thrombosis (DVT) of the lower limb is unreliable and individual symptoms and signs are of little value.\textsuperscript{2} Contrast venography has traditionally been the “gold standard” for the diagnosis of DVT but venography has been replaced in most centres by colour flow duplex ultrasonography. Venography and ultrasonography are comparatively expensive investigations and over 75\% of the investigated patients will prove not to have a DVT.\textsuperscript{3} In the occasional patient there may be impaired fibrinolytic activity.\textsuperscript{4} D-dimer concentrations and this has been shown in some studies.\textsuperscript{11, 12} In the occasional patient there may be impaired fibrinolytic activity.\textsuperscript{13} With the SimpliRED test there is a visual end point of agglutination of red blood cells and some studies have shown that interobserver variability in test results is high.\textsuperscript{14, 15} D-dimer tests generally have a high NPV but cannot be used in isolation as screening tests. D-dimer tests are therefore often used in conjunction with clinical probability scoring, plethysmography, or ultrasonography to reduce the need for further imaging.\textsuperscript{16–20} Although sensitive for venous thrombosis, high concentrations of D-dimer are insufficiently specific for making a positive diagnosis because they occur in other disorders such as malignancy, pregnancy, inflammatory conditions, and after surgery. There are four types of D-dimer assays commercially available—enzyme linked immunosorbent assays (ELISA), latex agglutination assays, a whole blood agglutination assay (SimpliRED), and an immunochromatography test (Simplify). The SimpliRED and Simplify tests are suitable for use as bedside tests whereas the ELISA and latex agglutination tests need to be performed in a laboratory. There are many quantitative latex agglutination and ELISA tests available\textsuperscript{21, 22} and the conventional ELISA is considered to be the gold standard for determination of D-dimer concentration. However, these laboratory based methods have the disadvantage that there can be a considerable delay in receiving the results in the emergency department.

**Abbreviations:** DVT, deep vein thrombosis; ELISA, enzyme linked immunosorbent assay; NPV, negative predictive value; PPV, positive predictive value
Moreover, the laboratory based methods are more suited to the assay of many samples in one batch rather than individual samples performed as soon as the patient attends the emergency department. A number of rapid D-dimer assays are available and include Tinaquant, Liatest, and Vidas D-dimer. These tests have sensitivities similar to conventional ELISA tests.

The SimpliRED D-dimer test has been used widely as a near patient test in emergency departments and provides a result within five minutes. However, the sensitivity of the SimpliRED test has been reported to vary from 61% to 100%.

A new near patient assay (Simplify D-dimer) has recently been introduced but there are currently no publications that have evaluated its efficacy. The aim of this study was to evaluate the use in the emergency department of the Simplify D-dimer test. The Simplify D-dimer test was compared with the (near patient) SimpliRED D-dimer assay and a (laboratory based) latex agglutination assay (Auto-D-dimer 700 on a Thromboscreen 400C analyser).

**METHODS**

The subjects were recruited into the study over a 21 month period starting in April 2001. The study was approved by the Bro-Taf Health Authority Local Research Ethics Committee and the subjects all gave informed written consent.

Patients presenting in the emergency department with clinical features suspicious of DVT were entered into the study, but the pre-test probability scores as performed in some studies were not calculated. Contrast venography was used to investigate DVT in all patients. As contrast venography is considered to be the gold standard, there was no follow up of patients with a negative venogram.

Patients were excluded from the study if they were less than 18 years old, had experienced recent trauma (<6 weeks), had undergone recent surgery (<6 weeks), were pregnant, had an underlying malignancy, or if they were having anticoagulant treatment. Patients were also excluded from the study if we were unable to perform venography (because of technical difficulties, or previous reaction to contrast).

Venous blood was obtained from each patient and a Simplify D-dimer test was performed immediately in the emergency department. Venous blood was also collected into plastic 4.5 ml Tri-sodium citrate (0.105M) Becton-Dickinson vacutainers in a ratio of 9:1 (blood:anticoagulant) and sent to the haematology laboratory for assessing with the SimpliRED and latex agglutination tests.

### Simplify D-dimer

The Simplify D-dimer assay consists of an individual test device in which the active ingredients are murine monoclonal antibody (DD3B6/22) specific for D-dimer conjugated to colloidal gold particles, a second D-dimer specific murine monoclonal antibody, and a sheep antimurine IgG antibody. Some 35 μl of whole blood is added to the sample well followed by two drops of buffer. The device was left lying flat for 10 minutes and then checked for a pink/purple line in the procedural control zone of the reading window. The absence of a line in this zone renders the test invalid. A positive result was indicated by the presence of a pink/purple line in the test zone of the reading window indicating a D-dimer concentration of greater than 80 ng/ml. A negative result was indicated by a complete absence of a line in the test zone of the reading window.

### SimpliRED D-dimer

The SimpliRED D-dimer tests were performed in the haematology laboratory on the citrated whole blood. The methodology for the SimpliRED D-dimer assay has been described elsewhere. The assay contains a bispecific antibody that reacts with high affinity to D-dimer (3B6/22), and with a red cell binding antibody (RAT-IC3/86). In the presence of D-dimer concentrations >120 ng/ml, the antibody induces agglutination of the patient’s red blood cells.

### Latex agglutination test

After centrifugation of the citrated venous blood at 2000–3000 g for 10 minutes, the plasma was then separated and frozen at −20°C for a maximum of one month. These samples were batch analysed for D-dimer using the Auto D-dimer (700) kit on the Thromboscreen 400C analyser.

The Auto D-dimer (700) is a quantitative latex micro-particle enhanced turbidimetric immunoassay that uses monoclonal antibody coated (MA-8D3) latex particles. In the presence of D-dimer, the particles aggregate thus increasing turbidity. The increase in scattered light is proportional to the amount of D-dimer in the sample.

The turbidity change in the sample is measured on a Thromboscreen 400C photometer that is set to measure at 705 nm. Photometric detection is started when the latex reagent is added to a cuvette containing the patient’s sample. The change in optical density (OD = Extinction) is monitored by the instrument. The turning point of the reaction curve is the final result and is displayed as ng/ml. The upper limit of the normal range was 120 ng/ml.

To avoid any bias, the haematology staff were unaware of the venogram results. The radiology staff performing the venograms were also blinded to the D-dimer results. The emergency department performed the Simplify D-dimer test before the venogram.

![Figure 1](http://www.emjonline.com/supplemental)
The sensitivity, specificity, positive predictive value (PPV), NPV, and 95% confidence intervals (CI) were calculated.

RESULTS
A total of 187 patients were enrolled into this study, 86 men and 101 women. Table 1 summarises the results and table 2 lists the sensitivity, specificity, NPV, and PPV for each of the D-dimer tests.

DISCUSSION
In this study 51 (27.2%) patients had a positive venogram and the Simplify D-dimer test proved to have the highest sensitivity of the three methods (table 2). The Simplify test “missed” three cases of DVT. In one of the cases (a 66 year old woman) with calf vein DVT and a false negative Simplify test both the latex agglutination and SimpliRED tests were also negative. Two male patients (aged 39 and 80) with proximal DVT also had a false negative Simplify test. In both these cases the SimpliRED and latex agglutination tests were both positive. The latex agglutination test had five false negative results and the SimpliRED test 13 false negative results.

With any new D-dimer test it is mandatory to have a high sensitivity and to have few false negative D-dimer results. With the SimpliRED test there is a visual end point of agglutination of red blood cells. It is much easier to interpret the Simplify test as the positive end point is a pink/purple line. It is expected that the interobserver variability will be less with the Simplify test than the SimpliRED test.

With quantitative D-dimer assays it is important to select a cut off level such that the D-dimer assay has a high sensitivity. However, the cut off level should not be so low that the specificity of the assay is also very low. A cut off level must be chosen such that there is a balance between sensitivity and specificity. The upper limit of the normal D-dimer range was initially chosen to be 120 ng/ml for the latex agglutination test (based on a normal cohort of laboratory staff). The five false negative D-dimer results were 70 ng/ml or less. Unless the cut off level was changed to less than 70 ng/ml the sensitivity of the assay would not be improved and there would be a very low specificity at a cut off level of 70 ng/ml.

It has been reported that there is a significantly lower sensitivity for the SimpliRED test if citrated whole blood is used instead of fingerstick capillary samples. Although the manufacturers of SimpliRED suggest that fingerstick capillary blood can be used, in the emergency department it is usually easier to take venous blood, as other blood investigations often need to be performed. In this study we used citrated venous blood for the SimpliRED test because the sample was sent to the haematology laboratory. The Simplify test was performed on venous blood.

The increased sensitivity of the Simplify D-dimer test when compared with the other two tests was at the expense of specificity (table 2). The specificity of the Simplify test was only 40.4%, compared with SimpliRED (83.1%) and latex agglutination (43.4%).

CONCLUSION
The Simplify D-dimer is easy to use, can be performed in 10 minutes, and the sensitivity was 94.1% in this study. This test is more suitable for use in the emergency department than the SimpliRED test. However, despite the high NPV of 94.8%, this test should be used in conjunction with clinical probability scoring, plethysmography, or ultrasonography to reach failure rates of less than 1%.

The Simplify test costs about £7, which is similar to most D-dimer tests. However, the laboratory based tests also need an analyser that entails a considerable initial capital cost. The cost of contrast for venography is on average over £30 and more importantly, ultrasonography and venography are very time consuming for skilled hospital staff. Despite the lack of specificity of the Simplify test, any reduction in further imaging would make the test worthwhile from an economic viewpoint.

CONTRIBUTORS
DN drafted the original manuscript, did the statistical analysis, and the laboratory investigations. CT initiated and coordinated the study, examined the subjects, and did the Simplify tests. AV was the radiology lead for the study and performed many of the venograms. SD did many of the laboratory investigations. KM was the haematology lead for the study. MO, VR, and AH examined the subjects and did the Simplify tests. All authors contributed to the final draft of the paper. CT is the guarantor.

A colour version of the figure is available on the journal web site (http://www.emjonline.com/supplemental).

Table 2 Sensitivity, specificity, NPV, and PPV of the investigated D-dimer assays

<table>
<thead>
<tr>
<th>Test</th>
<th>Sensitivity %</th>
<th>Specificity %</th>
<th>NPV %</th>
<th>PPV %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Simplify</td>
<td>94.1 (87.7 to 100)</td>
<td>40.4 (32.2 to 48.7)</td>
<td>94.8 (91.0 to 98.6)</td>
<td>37.2 (28.9 to 45.6)</td>
</tr>
<tr>
<td>SimpliRED</td>
<td>74.5 (62.5 to 86.5)</td>
<td>83.1 (76.8 to 89.4)</td>
<td>89.7 (82.0 to 97.3)</td>
<td>62.3 (50.1 to 74.5)</td>
</tr>
<tr>
<td>Auto D-dimer</td>
<td>90.2 (82.0 to 98.4)</td>
<td>43.4 (35.1 to 51.7)</td>
<td>92.2 (87.4 to 96.9)</td>
<td>37.4 (28.8 to 45.9)</td>
</tr>
</tbody>
</table>

The figures in parentheses are the 95% confidence intervals.

REFERENCES

www.emjonline.com
Dempfle CE, Turkstra F, 13 Eisenberg PR, Lennox AF, Mauron T, 15 Chapman CS, Kelly J, 8 Rudd A, Lewis RR, 11 Neale, Tovey, Vali, et al.

CONTINUING MEDICAL EDUCATION

ACCESS: the acute cerebral CT evaluation stroke study

Do you interpret CT brain scans within the first few hours of stroke? We run the acute cerebral CT evaluation stroke study (ACCESS), an internet based, interactive, CT reading tool designed to evaluate and improve CT reader reliability in detecting early infarct sign on CT. Readers log on to a web server (http://www.neuroimage.co.uk), complete a few details about background training and experience in viewing CT scans, view the study CT scans, and answer questions about each scan on the same screen. It is our intention to investigate the cross disciplinary recognition of stroke and to improve upon pre-existing paradigms in the diagnosis of acute stroke on CT.

There are 56 scans, reviewed in batches of about 10 scans each of which take about 20 minutes to complete. We will present the results of the study once it is completed later this year. Participation in ACCESS counts towards five continuing medical education category 1 credits in the UK. Accreditation was awarded by the Royal College of Radiologists, but because accreditation is not specific to RCR members, there is cross recognition between all UK Royal Colleges.

J Wardlaw

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